USE OF SYNCHROTRON RADIATION IN STUDIES OF PROTEIN STRUCTURE AND NUCLEIC ACID BINDING

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Interactions between proteins and DNA or RNA are not governed by simple lock-and-key mechanisms. Instead, the conformation of both the protein and the nucleic acid may be altered upon binding. In some cases, a defined three-dimensional structure non-existant in the apo state may be brought about by protein-nucleic-acid complex formation. In addition, a thermodynamic stabilization of the folded state may be observed. High-resolution crystal structures, accessible through synchrotronbased diffraction experiments, are indispensable for elucidating these structural principles which will be illustrated with three examples.

(i) Binding of Krueppel-like factor 4 (Klf4) to double-stranded (ds) DNA carrying the Klf4 target sequence leads to an induction of tertiary structure in the protein's zinc-finger domain. The spatial disposition of the three C2H2 zinc fingers of Klf4 is determined by a rigid dsDNA scaffold.

(ii) The plasmid RP4-encoded repressor KorB [1, 2] displays structural flexibility which facilitates the protein's functional cooperation with co-repressor KorA on RP4 promoters [3]. Flexible linkers between folded domains of KorB are important for its interaction with KorA over a fixed distance, but with variable geometry.

(iii) Cold shock domains interact with singlestranded (ss) DNA or RNA. The bacterial major cold shock proteins Bs-CspB [4, 5] and Bc-Csp [6, 7] bind ssDNA and ssRNA sequence-specifically and with high affinity to a conserved surface [8, 9]. The nucleic-acid single strands acquire a defined 3D structure upon protein binding. The same protein surface is used for ssDNA and ssRNA binding to the homologous coldshock domain of the human transcription factor YB-1. YB-1 assumes spectral characteristics of a folded protein and is conformationally stabilized upon binding to a single-stranded nucleic acid.

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